

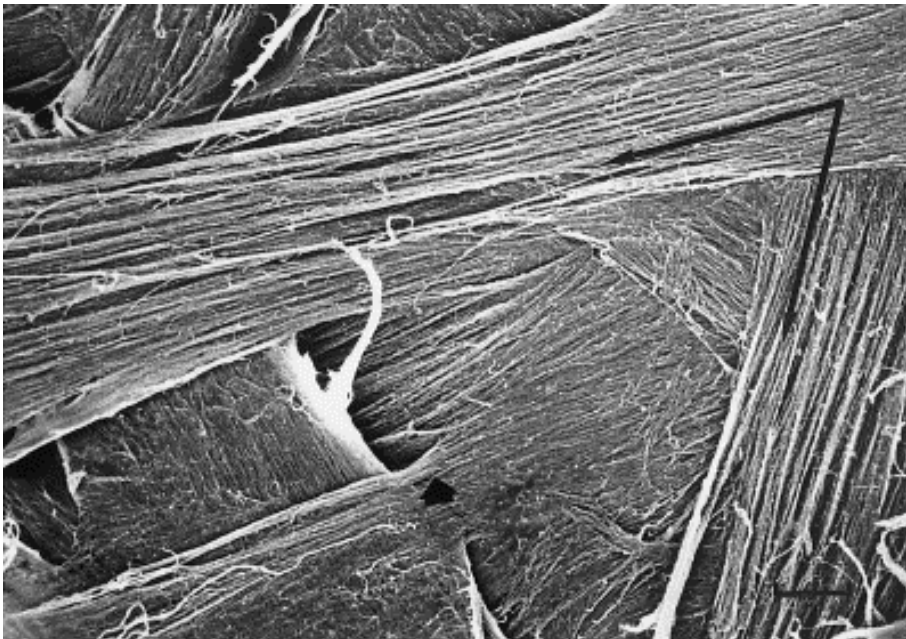


# Photothermal Chemistry of Collagen During Mid-IR Laser Ablation

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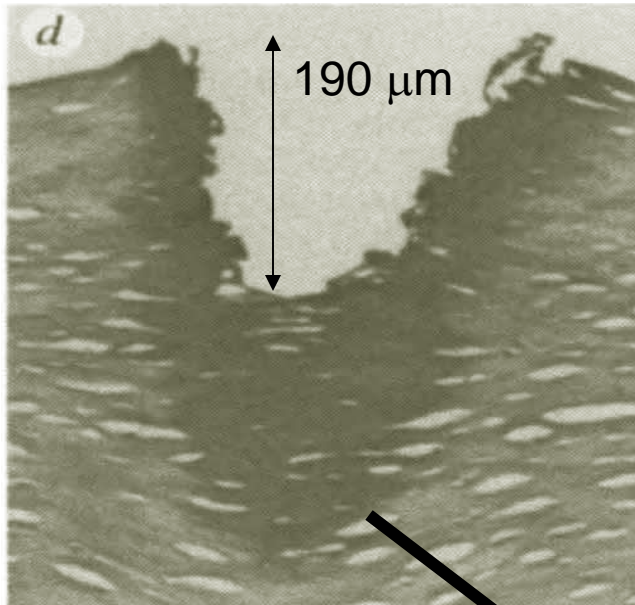


*Meek et al, 2001.*

To remove tissue with a laser, protein matrix must lose its structural integrity.

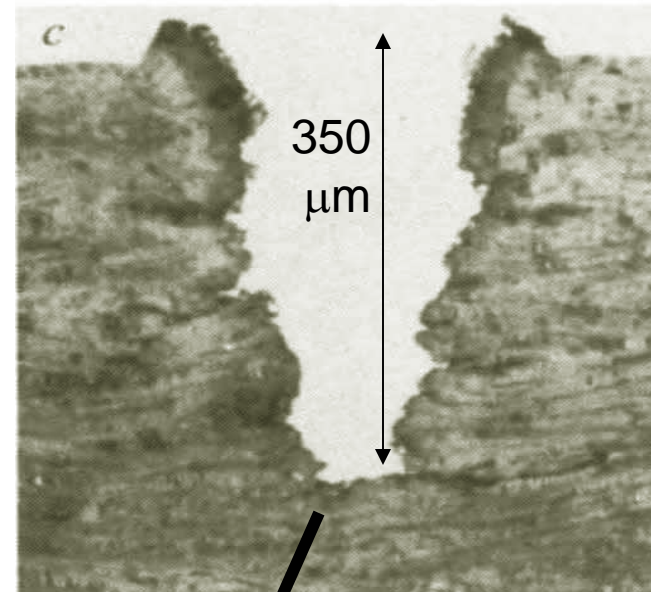
How does failure occur?

Can you influence it by your choice of laser parameters ( $\lambda$ ,  $\tau$ ,  $\Phi$ )?

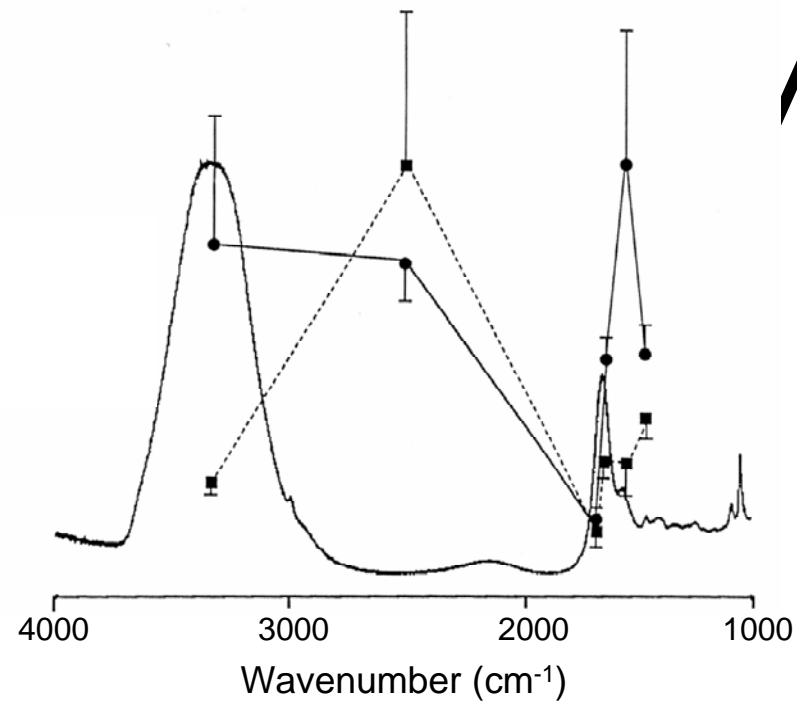


$\lambda = 3.0 \mu\text{m}$   
 $E_{\text{macro}} = 30 \text{ mJ}$

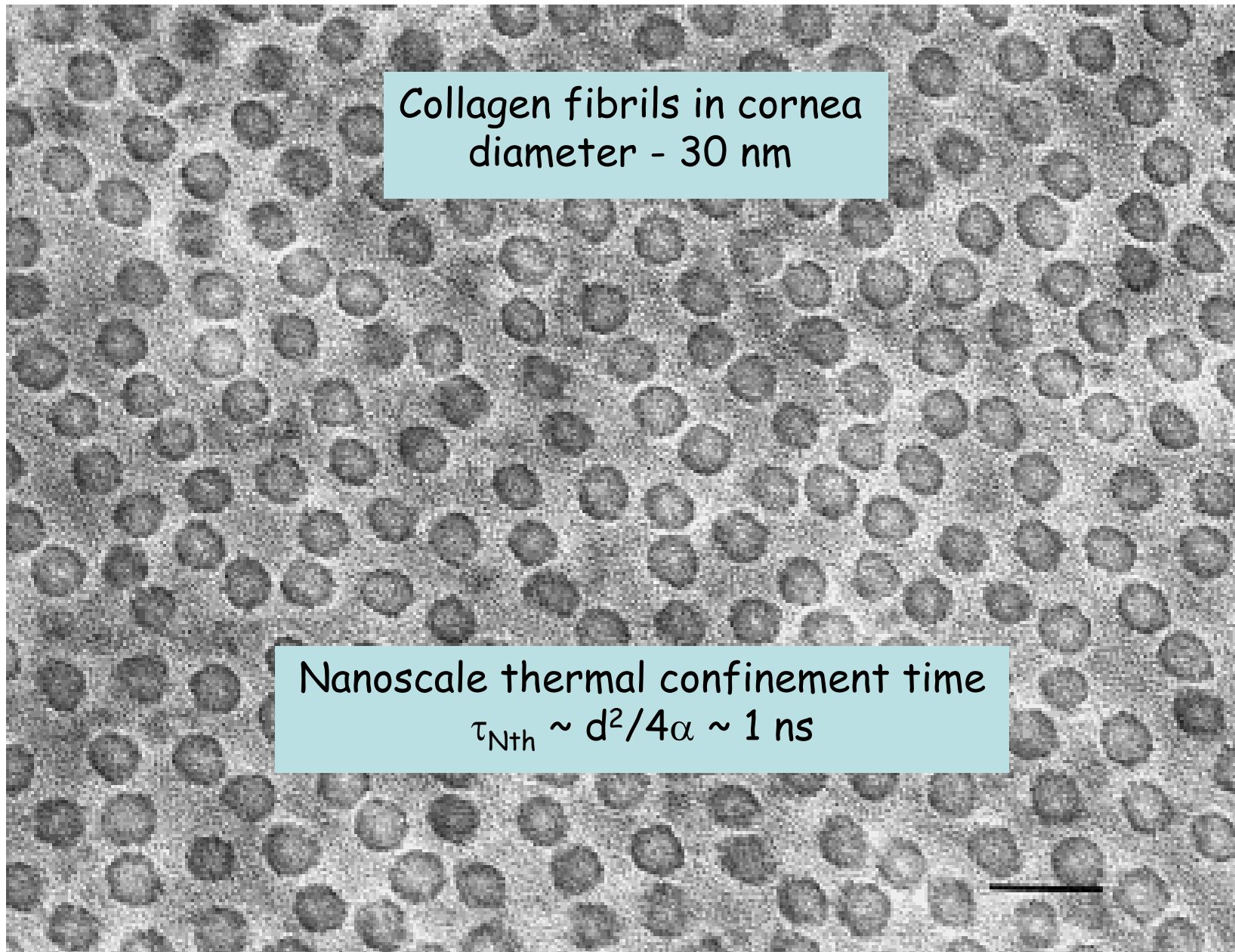
Corneas exposed  
to 10 macropulses  
at 4 Hz.



$\lambda = 6.45 \mu\text{m}$   
 $E_{\text{macro}} = 20 \text{ mJ}$



G. Edwards et al (1994)  
*Nature* 371: 416-419.



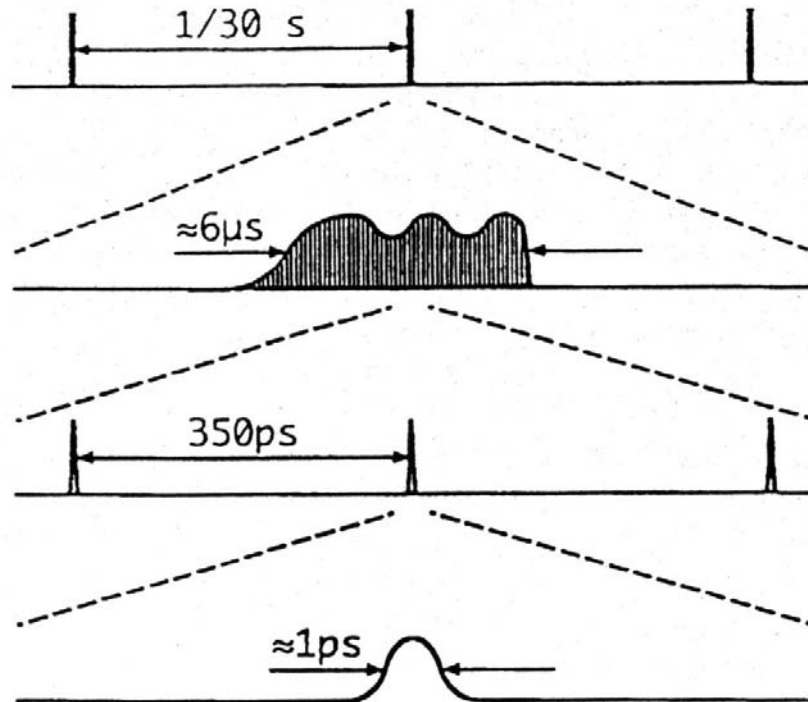
Collagen fibrils in cornea  
diameter - 30 nm

Nanoscale thermal confinement time  
 $\tau_{Nth} \sim d^2/4\alpha \sim 1 \text{ ns}$

*Meek et al, 2001.*



## Vanderbilt FEL Pulse Structure



$$P_{\text{avg}} = 0.6 \text{ W}, \quad I_{\text{avg}} = 7.2 \times 10^3 \text{ W/cm}^2$$

$$E_{\text{macro}} = 20 \text{ mJ},$$

$$\Phi_{\text{macro}} = 240 \text{ J/cm}^2$$

$$P_{\text{macro}} = 3.3 \text{ kW}, \quad I_{\text{macro}} = 4.0 \times 10^7 \text{ W/cm}^2$$

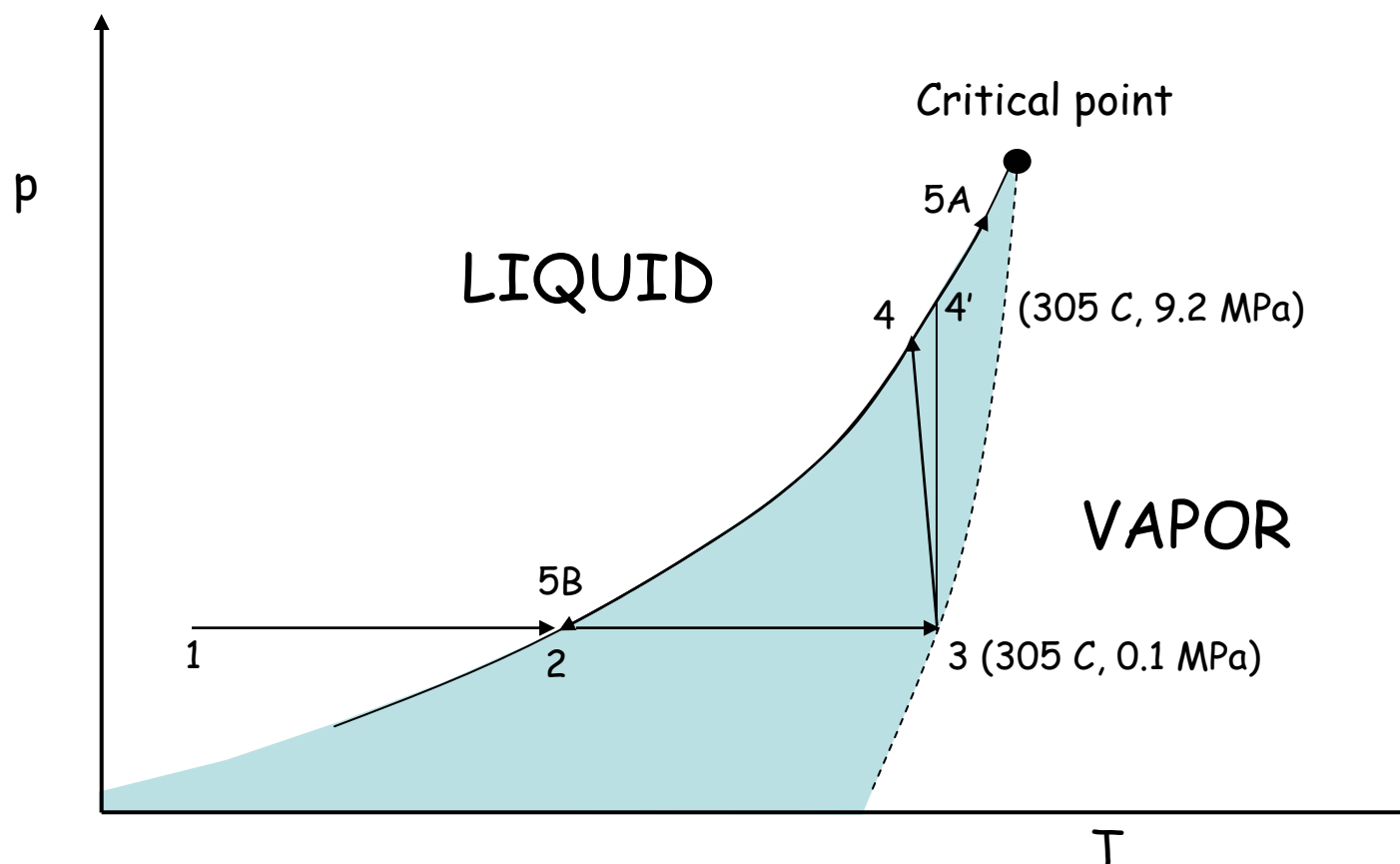
$$E_{\text{ps}} = 1.2 \mu\text{J},$$

$$\Phi_{\text{ps}} = 0.014 \text{ J/cm}^2$$

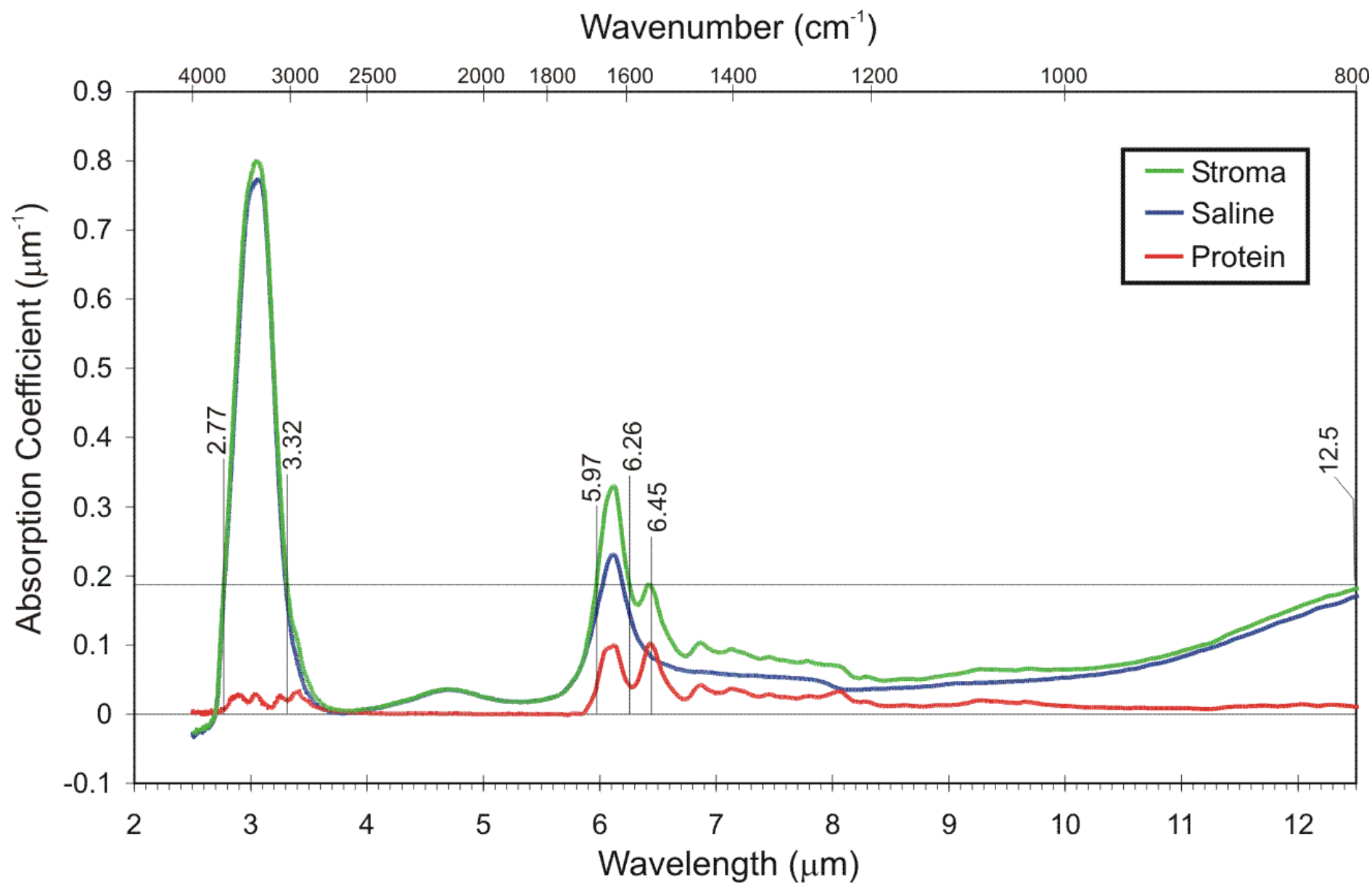
$$P_{\text{ps}} = 1.2 \text{ MW}, \quad I_{\text{ps}} = 1.4 \times 10^{10} \text{ W/cm}^2$$

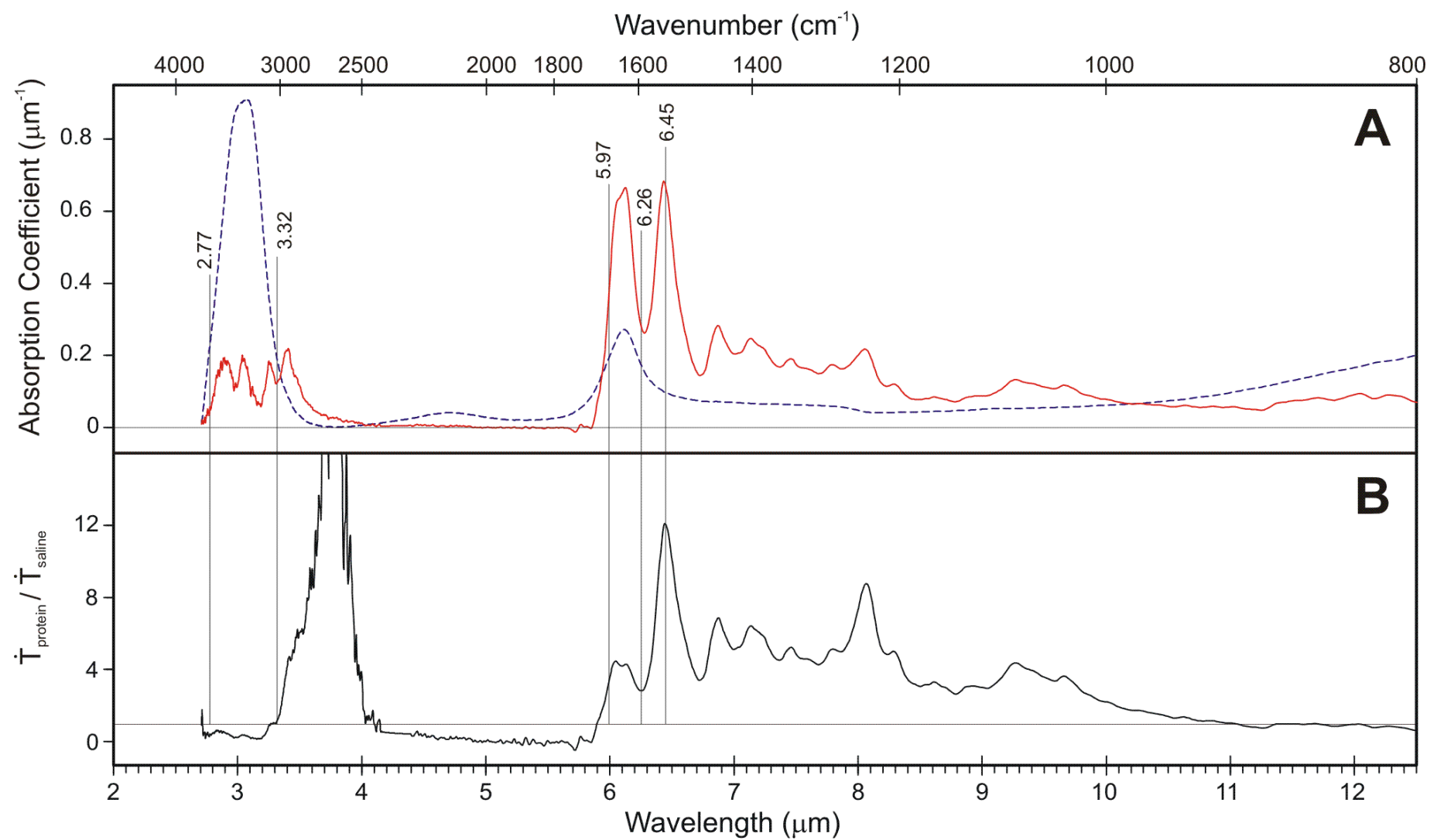


# Explosive Vaporization



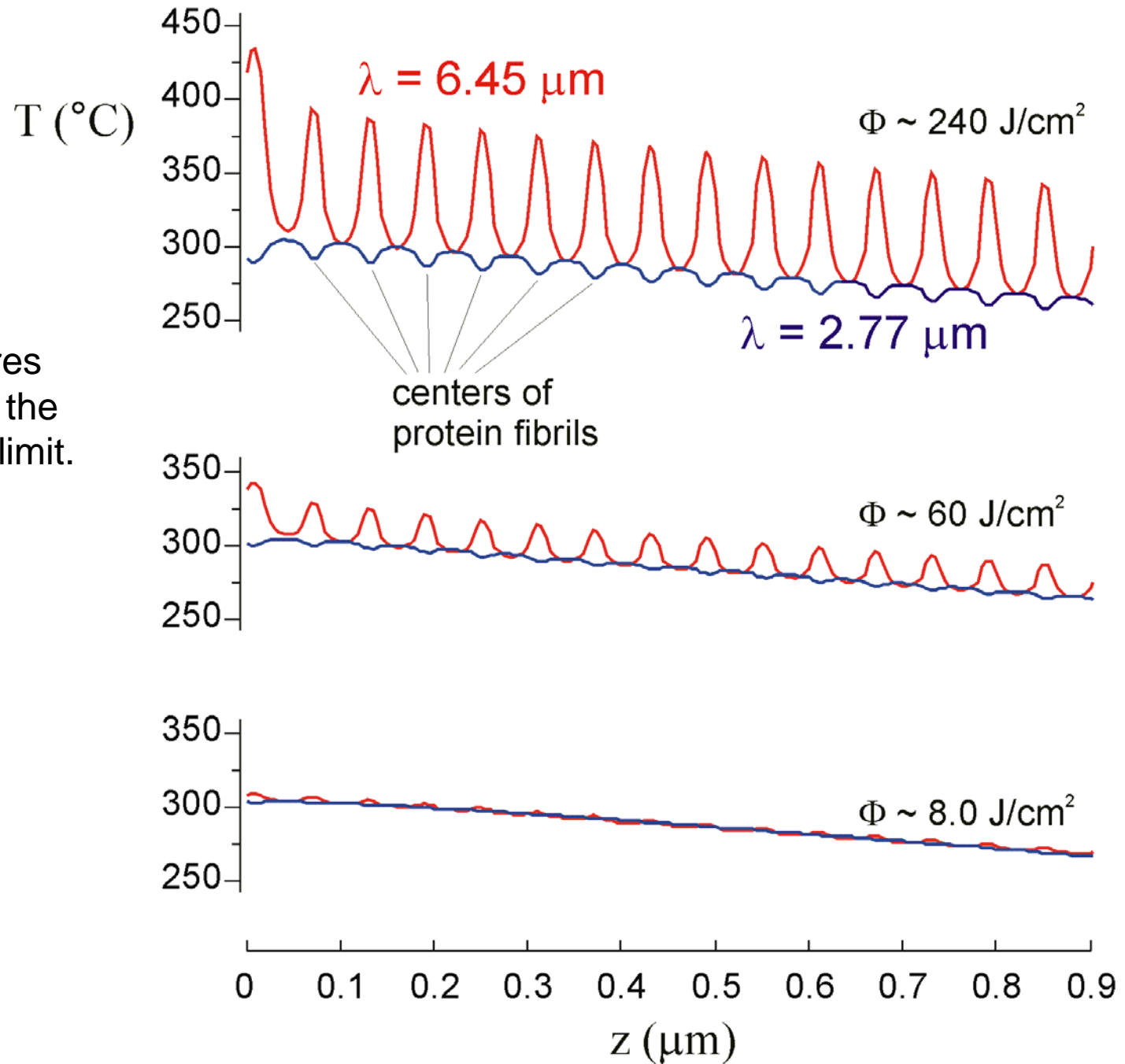
A. Vogel and V. Venugopalan (2003), Chem. Rev. 103: 577-644.







Predicted  
temperatures  
attained at the  
superheat limit.



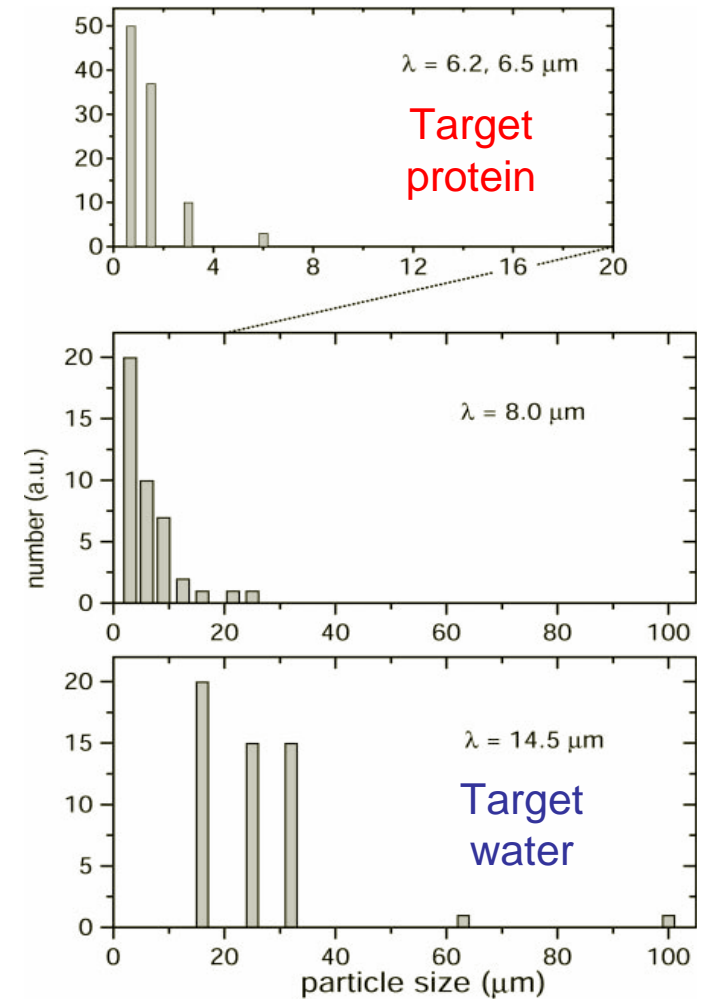
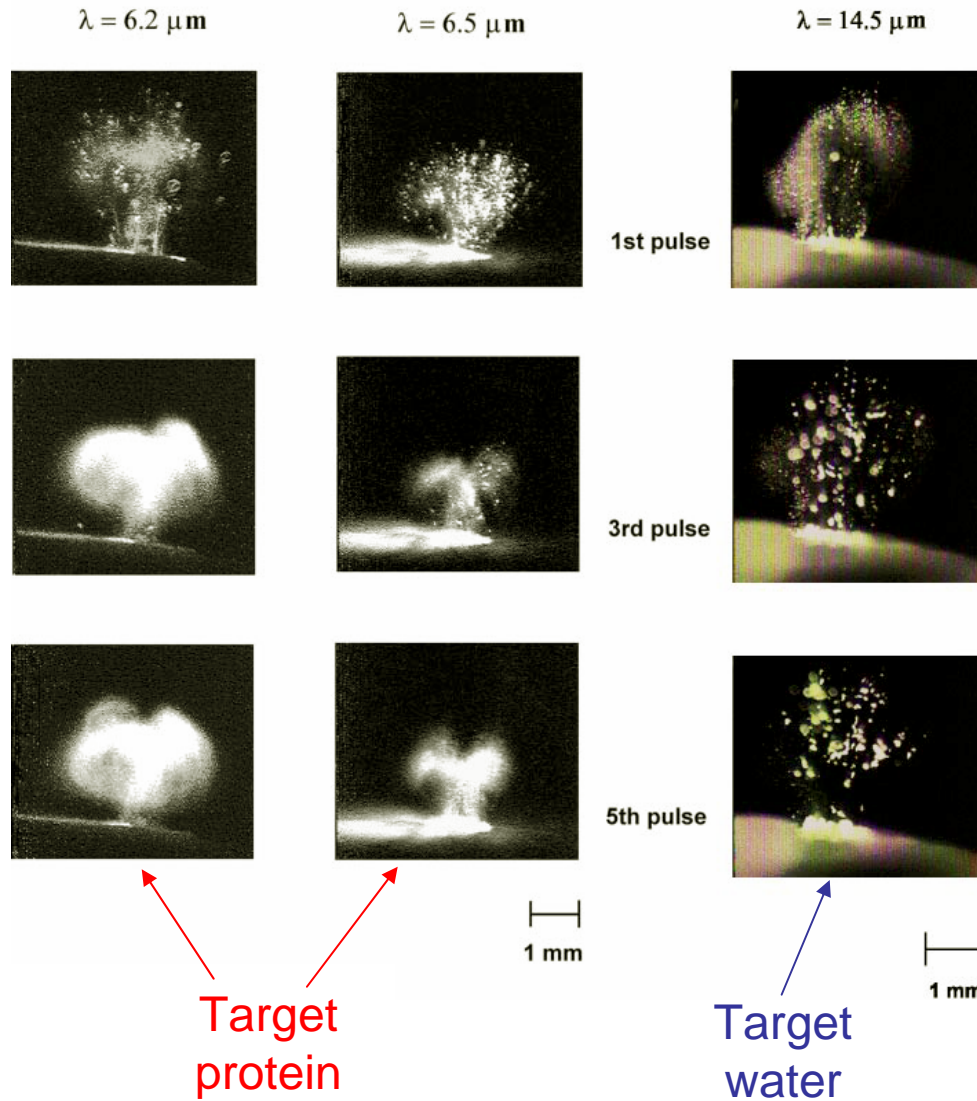


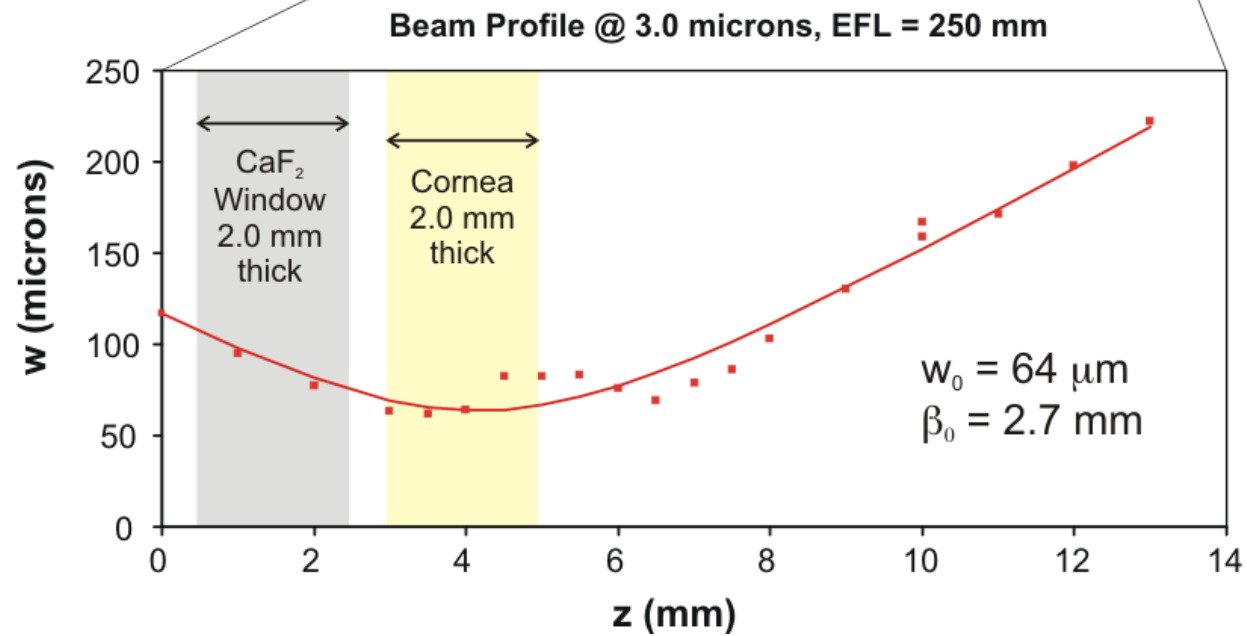
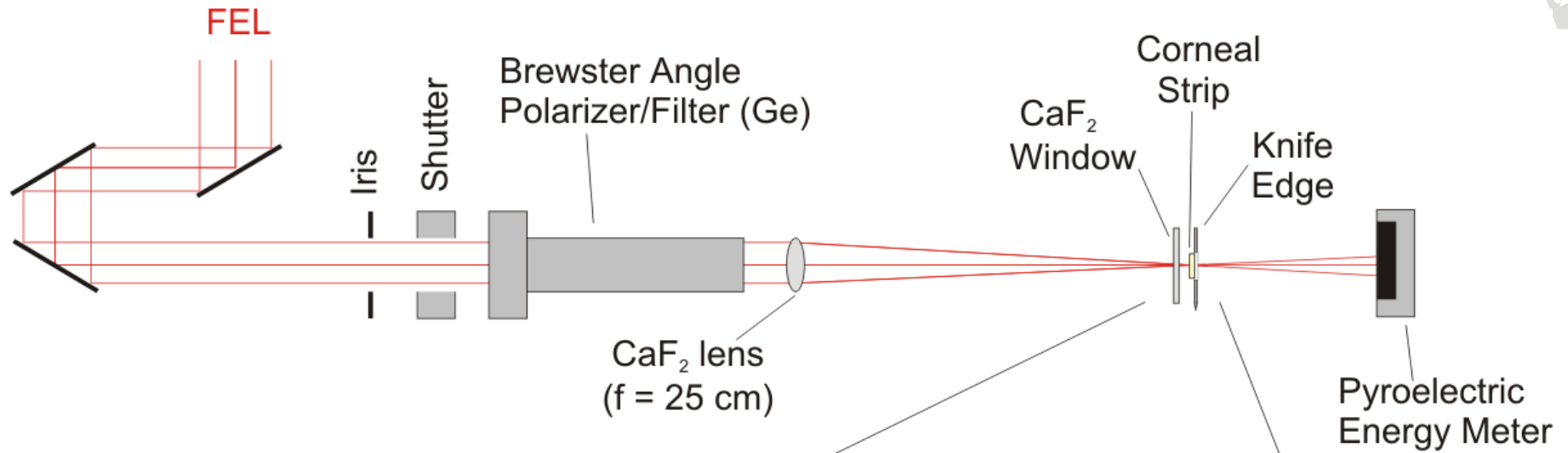


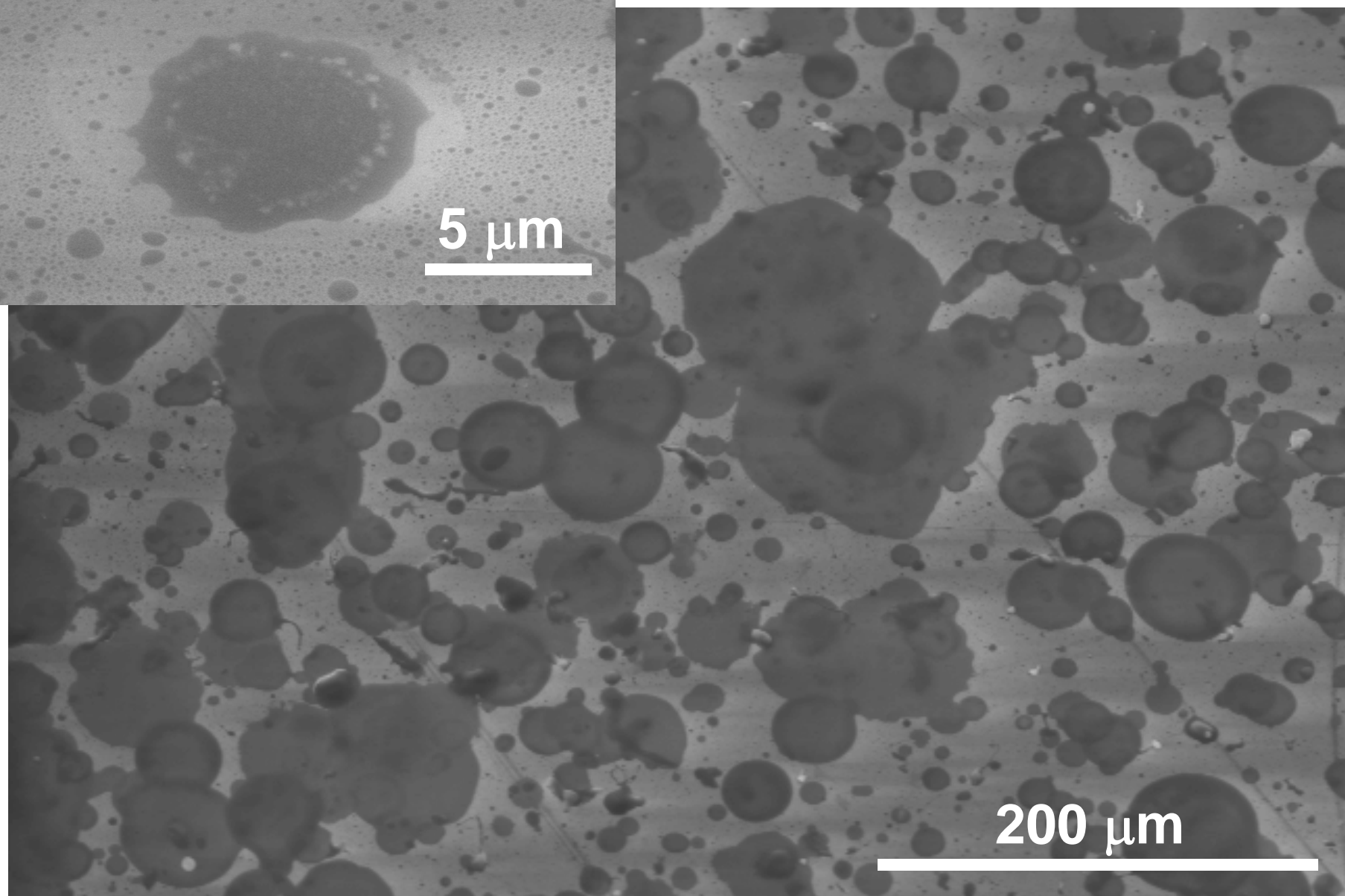
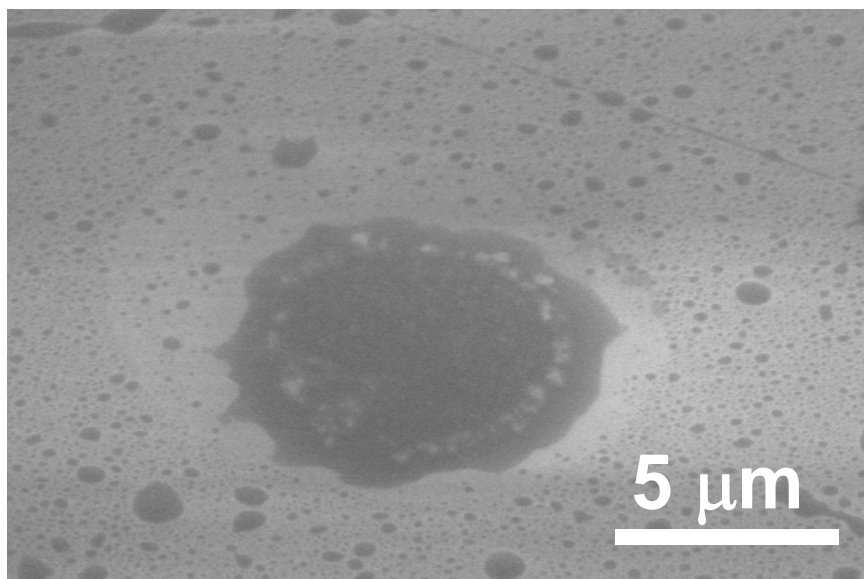
## Data from FELIX

J.M. Auerhammer et al (1999) *Appl. Phys. B* 68: 111–119.

$$E_0 = 20 \text{ mJ}, \delta t = 5 \mu\text{s}$$



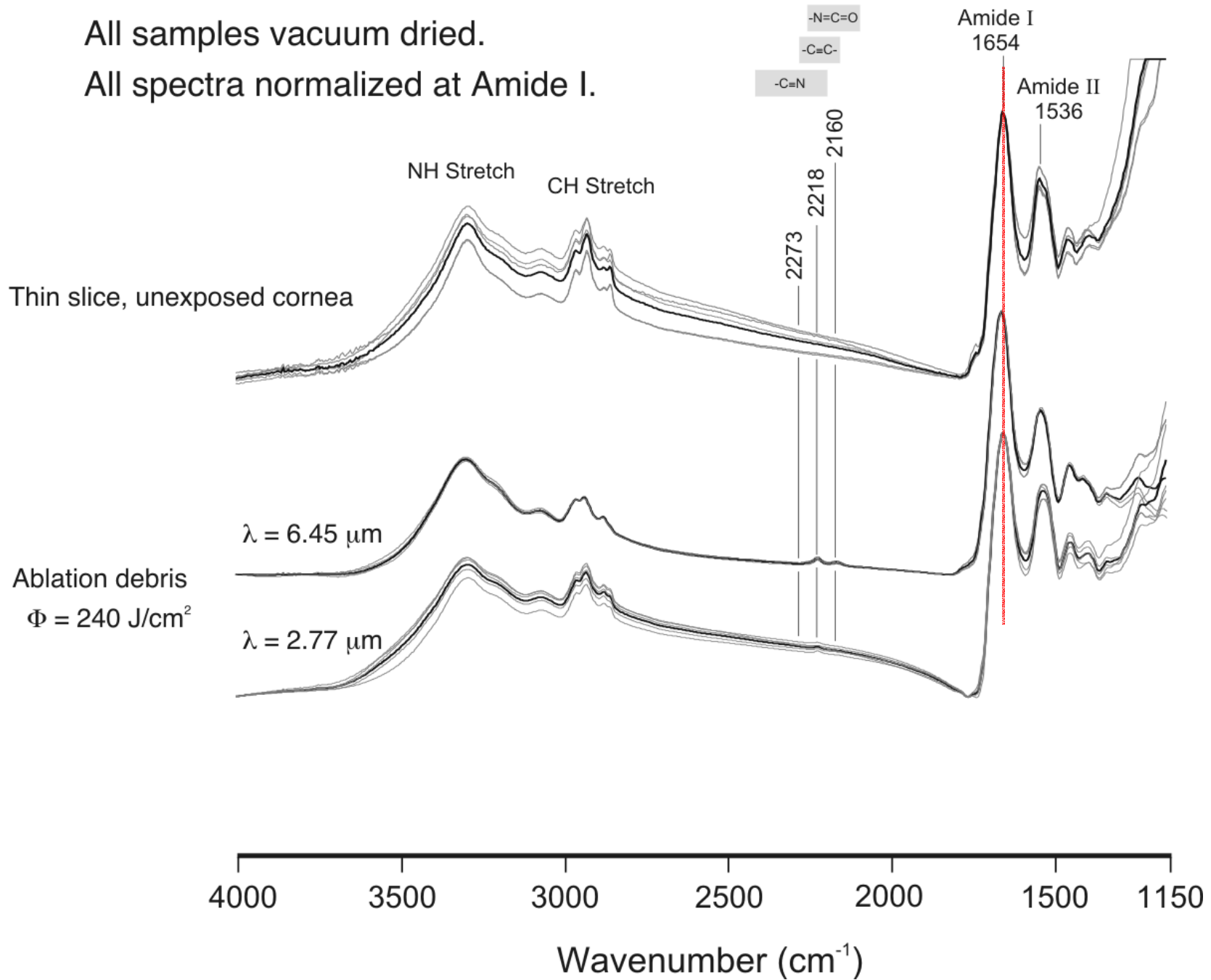






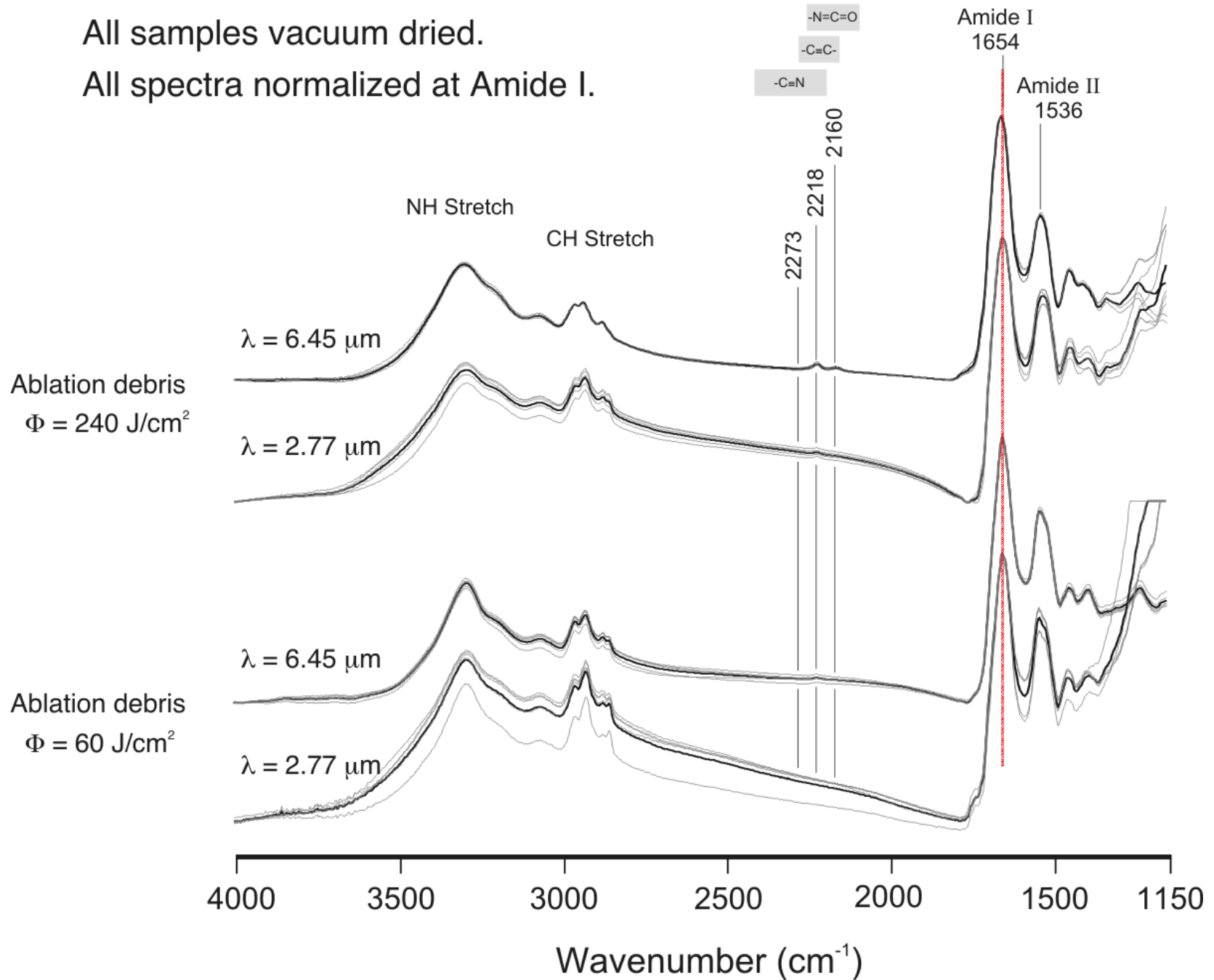
All samples vacuum dried.

All spectra normalized at Amide I.



All samples vacuum dried.

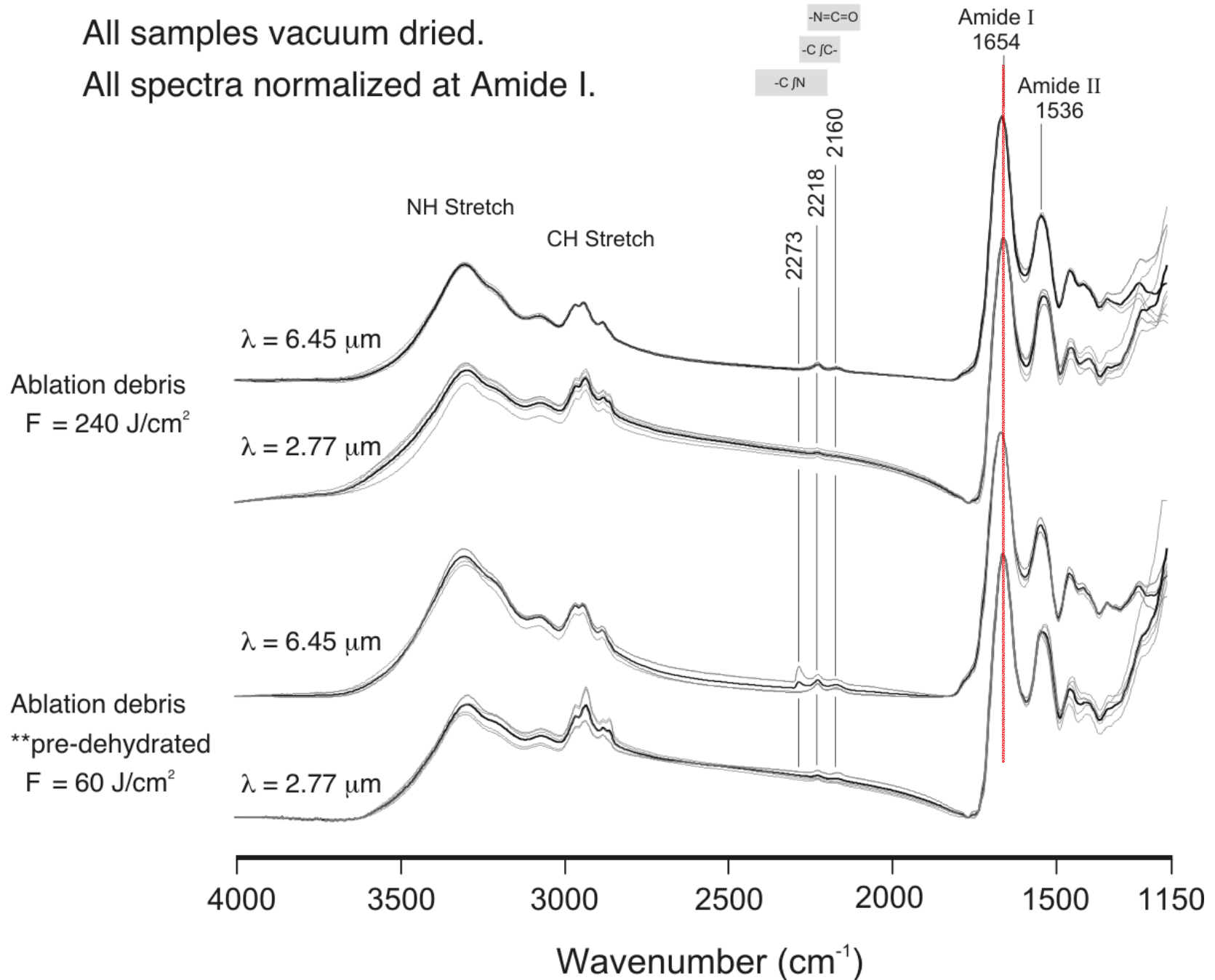
All spectra normalized at Amide I.





All samples vacuum dried.

All spectra normalized at Amide I.

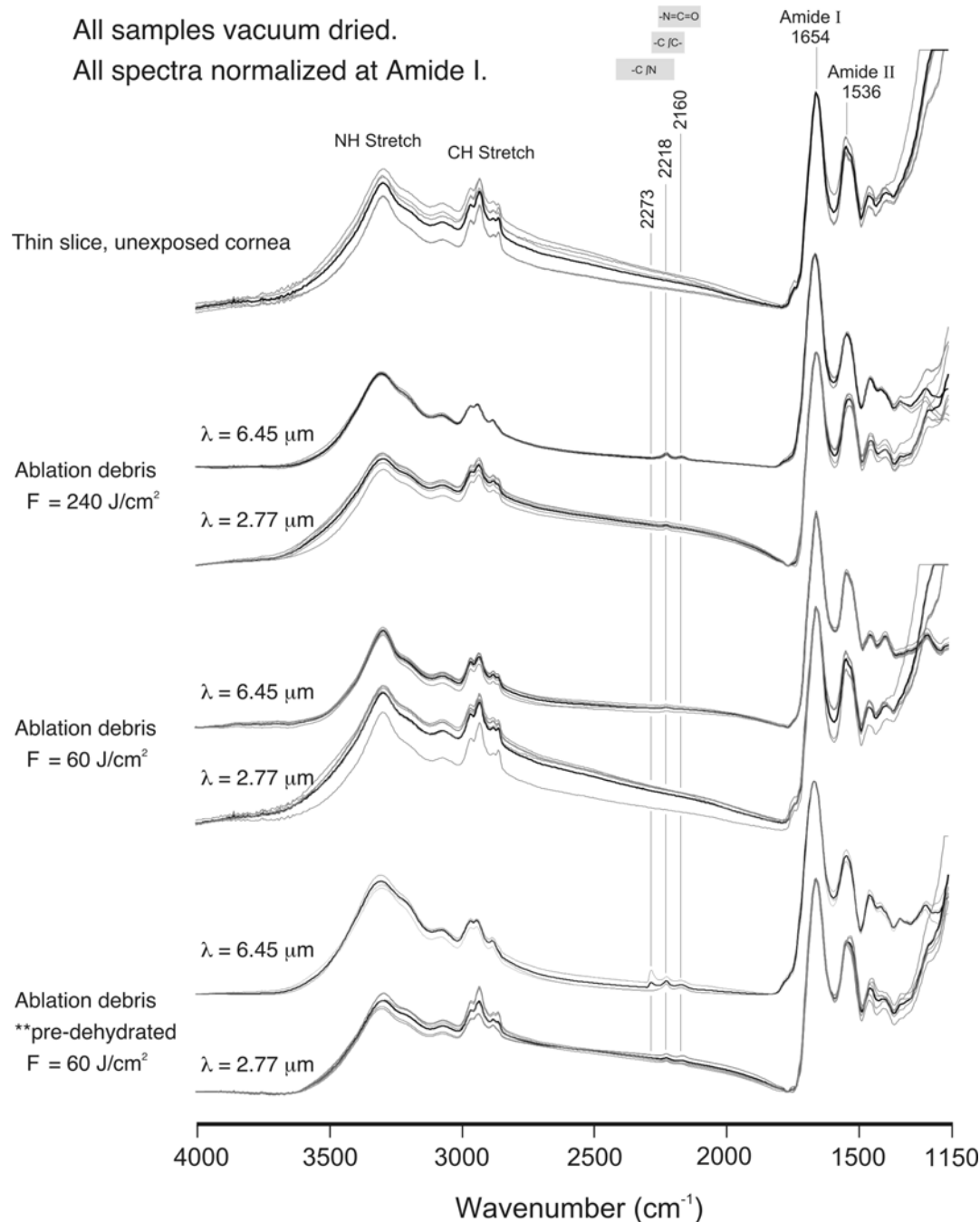






All samples vacuum dried.

All spectra normalized at Amide I.



Amide I frequency is NOT indicative of unfolded protein. For most samples it matches that of dry collagen triple helix. For others it is slightly upshifted.

Similar  $\sim 2200 \text{ cm}^{-1}$  bands observed on the surface of ablated bone, dentin and enamel. Previously assigned to cyanate ions.

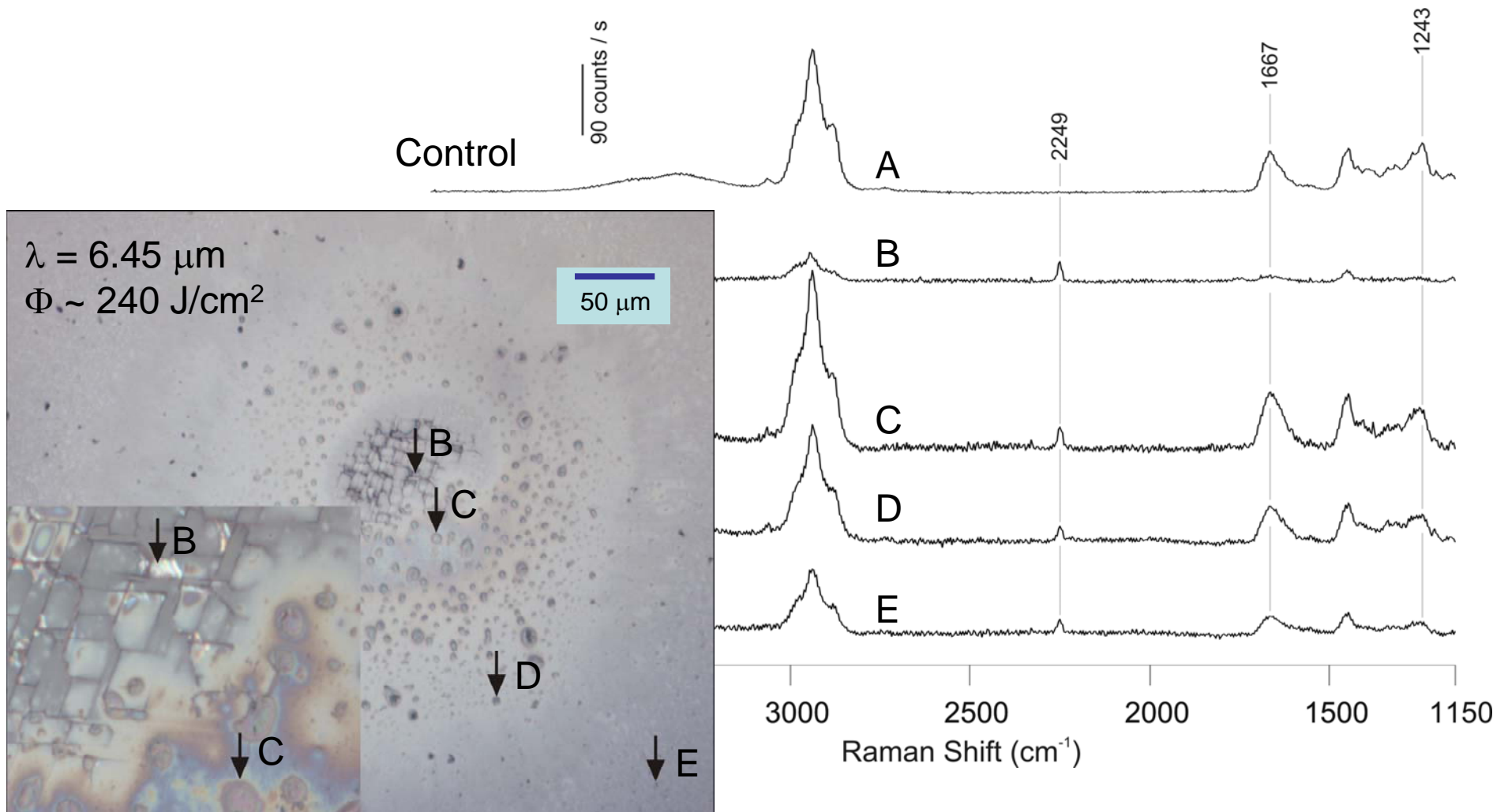
- Dowker and Elliott, 1979, 1983; Spencer et al, 1996.

Similar bands arise in thermally treated dentin ( $>300^\circ\text{C}$ ) and are accompanied by an ESR signal.

- Bachmann et al 2004.



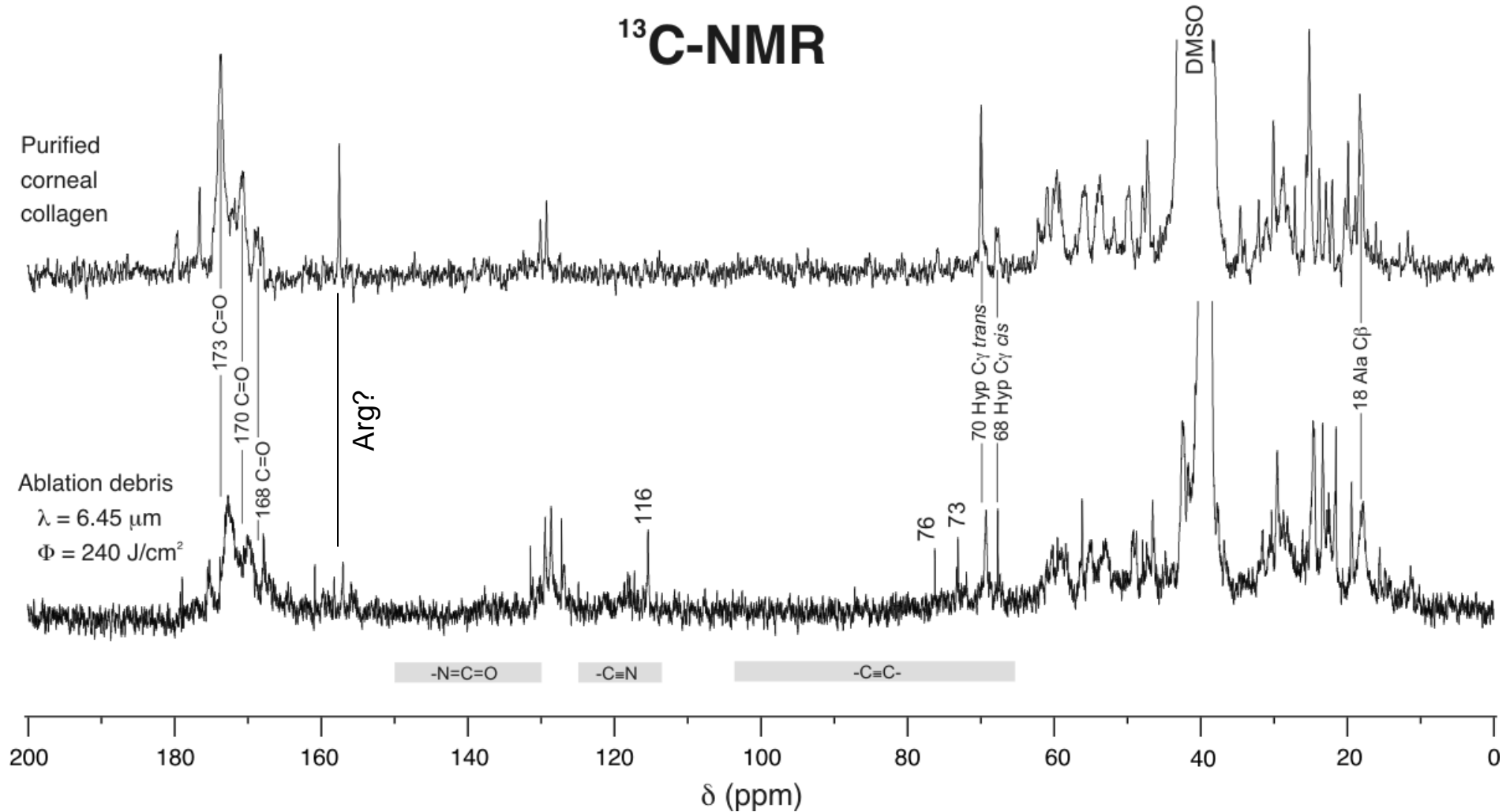
## Micro-Raman Spectra from SINGLE Macropulse Ablation







# <sup>13</sup>C-NMR



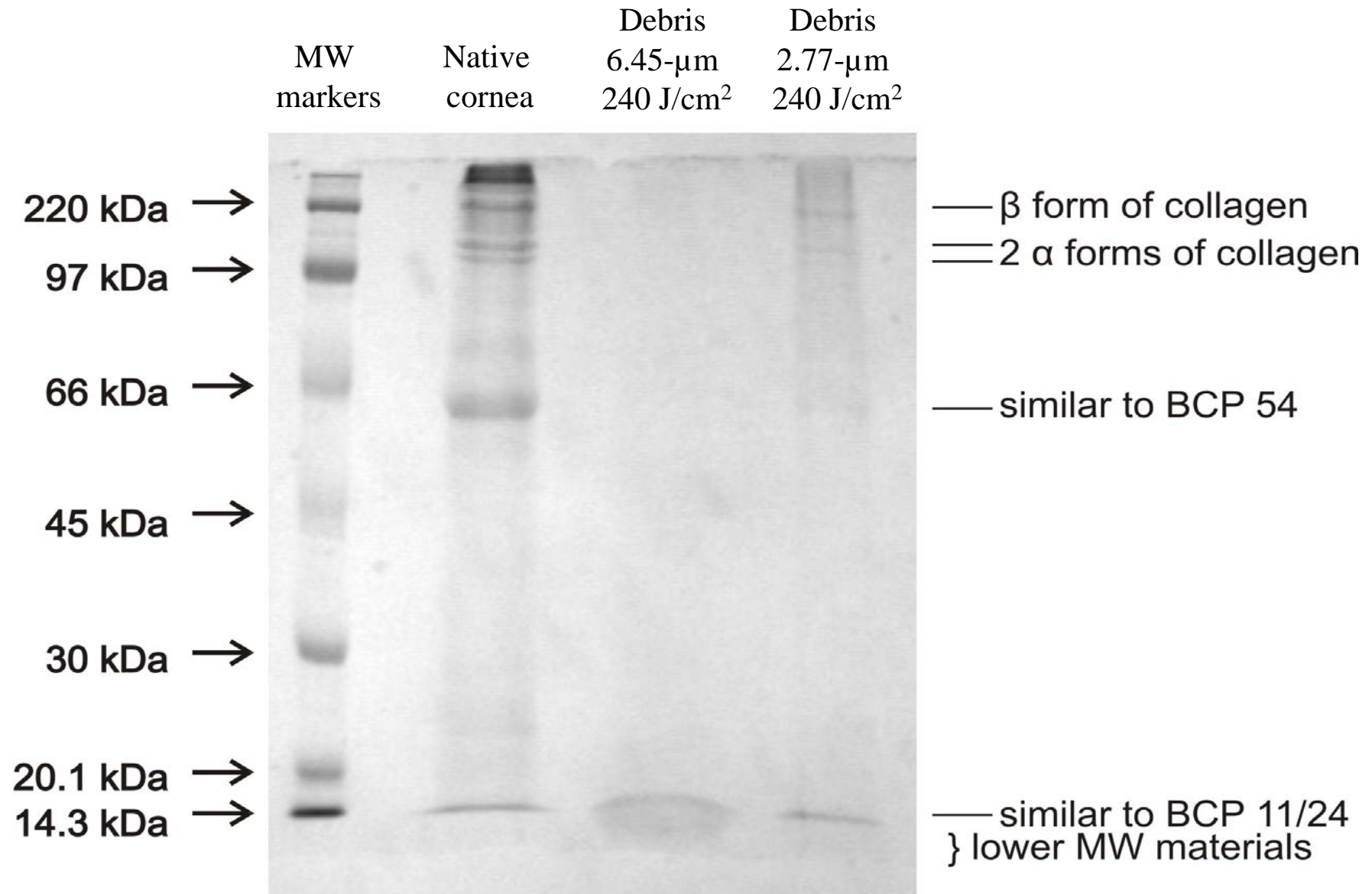
\*Purified collagen was prepared using limited *pepsin A* digestion of porcine cornea.

\*\*Ablation debris from ~200 CaF<sub>2</sub> windows was pooled.

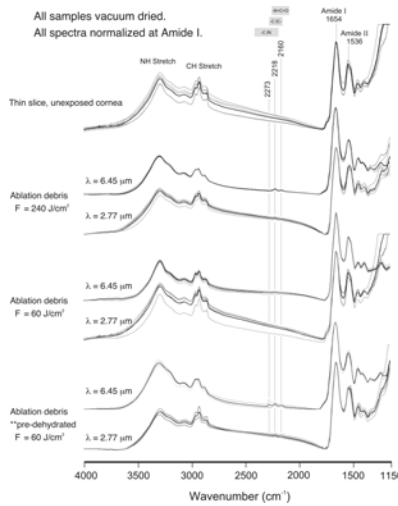
Band assignments from Saitô et al, 1984; Sarkar et al, 1984; Fujisawa et al, 1990; Saitô and Yokoi, 1992; Reichert et al, 2004



## SDS (5%) extracts of:

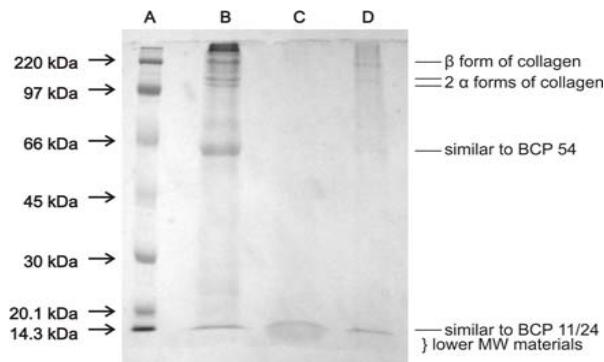
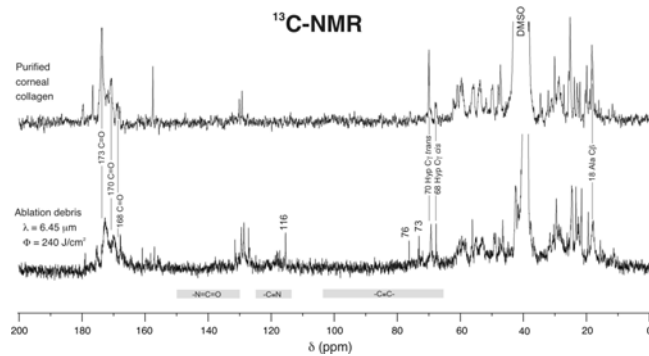


*Band assignments from Cooper et al, 1990;  
Bakker et al 1992.*



In the ablation debris, most of the primary and secondary structure of collagen remains intact.

But new chemical species (2-3) are also present. The combination of FTIR and  $^{13}\text{C}$ -NMR definitively assigns one of the new species as a nitrile group ( $-\text{C}\equiv\text{N}$ ) and suggests that the others could be alkynes ( $-\text{C}\equiv\text{C}-$ ). The formation of these species is wavelength-, fluence- and hydration-dependent.



SDS-PAGE reveals that collagen is fragmented into smaller peptides in a  $\lambda$ -dependent manner. (~10 breaks per 1000 residue protein chain for  $\lambda = 6.45 \mu\text{m}$ )